

RUMBRIN, A NEW CYTOPROTECTIVE SUBSTANCE
PRODUCED BY *Auxarthron umbrinum*

II. PHYSICO-CHEMICAL PROPERTIES AND
STRUCTURE DETERMINATION

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The structure of rumbrin (Fig. 1), a new cytoprotective substance, was elucidated by NMR spectral analysis. Rumbrin was found to possess a novel skeleton containing α -pyrone, tetraene and pyrrole moieties.

In the preceding paper¹⁾, we described the fermentation, isolation and biological activities of rumbrin, which is produced by *Auxarthron umbrinum*. This paper describes the physico-chemical properties and structural studies on rumbrin.

Rumbrin (**1**) is a red crystalline solid with the properties listed in Table 1. The molecular formula of **1** was determined to be $C_{20}H_{20}NO_3Cl$ by HRFAB-MS ($(M+H)^+$ m/z calcd: 358.1210, found: 358.1206).

Fig. 1. The total structure of rumbrin.

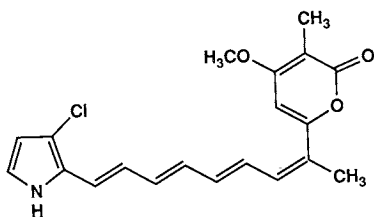


Table 1. Physico-chemical properties of rumbrin.

Appearance	Red needles
MP (dec.)	170~171°C
Molecular formula	$C_{20}H_{20}NO_3Cl$
HRFAB-MS Calcd:	358.1210
	Found: 358.1206 ($M+H$) ⁺
UV λ_{max} nm (ϵ)	269 (12,750), 341 (17,350), (in MeOH) 442 (33,800)
IR ν (KBr) cm^{-1}	3420, 2920, 1660, 1620, 1525, 1240
Rf value ^a	0.46
Soluble	MeOH, DMSO, $CHCl_3$, EtOAc
Insoluble	H_2O , hexane

^a Solvent system: $CHCl_3$ - MeOH (50:1) Kieselgel 60 F_{254} .

Fig. 2. 1H NMR spectrum of rumbrin in $DMSO-d_6$ (500 MHz).

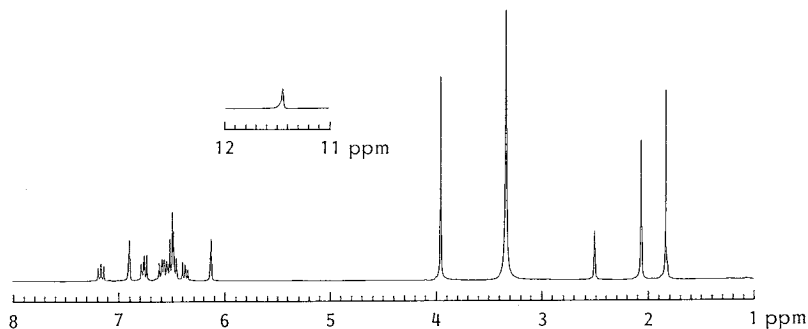


Table 2. The 500 MHz ^1H NMR and 125 MHz ^{13}C NMR spectral data for rumbrin^a.

Position	δ_{H}	δ_{C}	Position	δ_{H}	δ_{C}
1-NH	11.44 (dd 2.3, 2.6) ^b		12	6.47 (d 12.5)	134.7 (d)
2	6.90 (dd 2.6, 3.0)	120.2 (d)	13		124.8 (s)
3	6.13 (dd 2.3, 3.0)	109.0 (d)	14		159.1 (s)
4		111.5 (s)	15	6.50 (s)	96.2 (d)
5		125.9 (s)	16		165.7 (s)
6	6.49 (d 15.0)	120.2 (d)	17		100.3 (s)
7	6.75 (dd 11.0, 15.0)	124.8 (d)	18		163.4 (s)
8	6.59 (dd 11.0, 14.5)	135.9 (d)	19	2.05 (s)	21.1 (q)
9	6.37 (dd 11.3, 14.5)	131.3 (d)	20	3.95 (s)	56.6 (q)
10	6.54 (dd 11.3, 14.0)	137.9 (d)	21	1.92 (s)	8.5 (q)
11	7.16 (dd 12.5, 14.0)	128.9 (d)			

^a Taken in DMSO- d_6 .

^b Coupling constants in $J = \text{Hz}$.

The ^1H NMR spectrum of **1** (Fig. 2) showed 14 signals, which were attributed to two singlet CH_3 (δ_{H} 1.92 and 2.05), one OCH_3 (δ_{H} 3.95), one imine (δ_{H} 11.44) and 10 olefinic methine protons.

The ^{13}C NMR spectrum of **1** showed signals for 20 carbons. The distortionless enhancement by polarization transfer (DEPT) experiment assigned them to 3 methyl, 10 sp^2 methine, and 7 quaternary carbons including one ester carbonyl carbon (C-18) and 2 oxygenated sp^2 carbons (C-14 and C-16).

The ^1H - ^1H COSY spectrum established a tetraene structure composed of C-6~C-12 with *E* geometrical configurations for C-6, 8 and 10 apparent from the coupling constants ($J_{6,7} = 15.0 \text{ Hz}$, $J_{8,9} = 14.5 \text{ Hz}$ and $J_{10,11} = 14.0 \text{ Hz}$).

The HMBC experiment on **1**²⁾ showed long range couplings of 19- CH_3 to C-13 (δ_{C} 124.8) and C-14 (δ_{C} 159.1), 21- CH_3 to C-16 (δ_{C} 165.7), C-17 (δ_{C} 100.3) and C-18 (δ_{C} 163.4), 20- OCH_3 to C-16 and 15-H to C-13, C-14, C-16 and C-17. These correlations established the connectivities of C-13~C-18. Taking into consideration the number of oxygen atoms contained in **1** and the chemical shifts of C-14 and C-18, one oxygen atom must be inserted between C-14 and C-18. Thus, the existence of an α -pyrone unit in **1** was confirmed as shown in Fig. 3.

The HMBC experiment also showed long range couplings of 19- CH_3 to C-12 (δ_{C} 134.7), C-13 (δ_{C} 124.8) and C-14. Thus, the tetraene and the α -pyrone units are linked through C-13 (Fig. 3). The diagnostic ^{13}C chemical shift for C-19 (δ_{C} 21.1)³⁾ and NOE between 12-H and 19- CH_3 defined the configuration of the C-12~C-13 double bond as *Z*.

In the ^1H - ^1H COSY spectrum, cross peaks were observed among the two methine protons (2-H (δ_{H} 6.90, $J = 2.6, 3.0 \text{ Hz}$) and 3-H (δ_{H} 6.13, $J = 2.3, 3.0 \text{ Hz}$)) and an imine proton (δ_{H} 11.44, $J = 2.3, 2.6 \text{ Hz}$). In addition, long range couplings were observed from 3-H to C-2 (δ_{C} 120.2) and C-4 (δ_{C} 111.5), 2-H to C-3 (δ_{C} 109.0), C-4 and C-5 (δ_{C} 125.9), 6-H to C-4 and C-5, and 7-H to C-5 in the HMBC experiment.

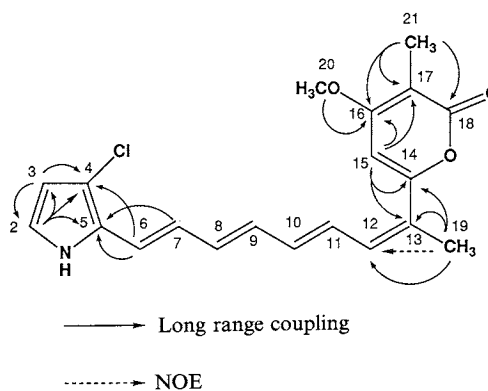
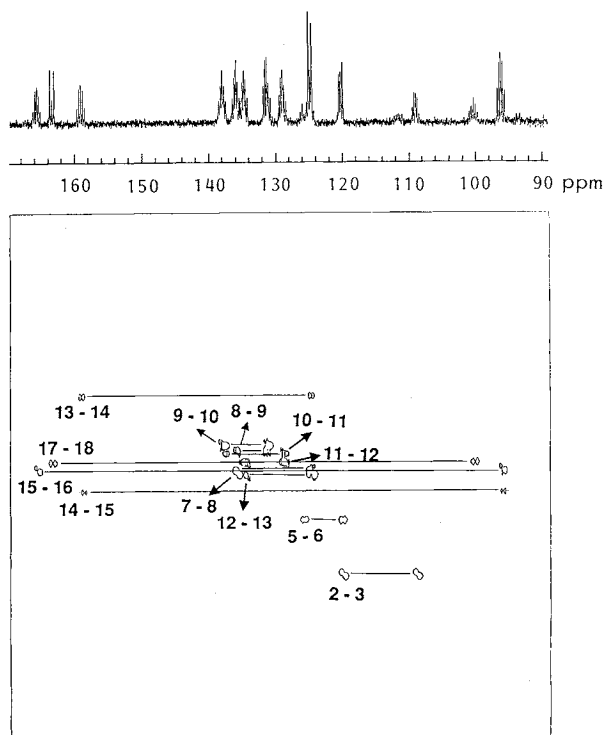
Fig. 3. ^1H - ^{13}C long range couplings and NOE.

Fig. 4. 2D INADEQUATE spectrum of [1,2- $^{13}\text{C}_2$]acetate labeled rumbrin.

These couplings indicated the presence of a 2,3-disubstituted pyrrole ring consisting of C-2~C-5, and the linkage to the tetraene unit at C-5. Therefore, attachment of the chlorine atom to the quaternary carbon C-4 was assigned (Fig. 3). Based on all these findings, the total structure of **1** was established to be (1*Z*,3*E*,5*E*,7*E*)-6-(8-(3-chloro-1*H*-pyrrol-2-yl)-1-methyl-1,3,5,7-octatetraenyl)-4-methoxy-3-methyl-2*H*-pyran-2-one (Fig. 1).

In the ^1H - ^{13}C COSY spectrum, the olefinic carbon signals could not be unambiguously assigned because of the overlapping of their proton signals. Therefore, we assigned these carbons and confirmed the structure of **1** using $^1J_{\text{C}-\text{C}}$ information. The biosynthetic origin of the polyene and α -pyrone units was expected to be mainly acetate⁴). Thus, an incorporation experiment with [1,2- $^{13}\text{C}_2$]acetate was carried out with a culture of *A. umbrinum* n13. By adding 1 g of sodium [1,2- $^{13}\text{C}_2$]acetate 48 hours after the beginning of the 1 liter culture, 4 mg of labeled **1** was obtained. A 2D INADEQUATE experiment using this sample confirmed the total structure of **1** and the assignments of all sp^2 carbons (Fig. 4).

Experimental

General

Mass spectra were measured on a VG Analytical ZAB-HF in the FAB mode using a *meta*-nitrobenzylalcohol matrix. UV spectra were recorded using a Hitachi U-3200 spectrophotometer. NMR spectra were obtained on a JEOL JNM-GX500 spectrometer with ^1H NMR at 500 MHz and ^{13}C NMR at 125 MHz. Chemical shifts are given in ppm using TMS as internal standard.

Labeled Compound

Sodium [1,2- $^{13}\text{C}_2$]acetate was purchased from MSD Isotopes.

Incorporation Studies

The incorporation studies were conducted as follows: The strain n13 was inoculated into 100 ml of a seed medium consisting of glucose 2.0%, peptone 0.5%, KH_2PO_4 0.5%, $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.05%, $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ 0.05% and agar 0.1% in a 500-ml Erlenmeyer flask (pH 6.0), and cultured at 25°C for 5 days on a rotary shaker (180 rpm). Three ml of this seed culture was inoculated into 100 ml of the production medium having the same composition as the seed medium in 500-ml Erlenmeyer flasks. After incubation at 25°C for 2 days on the rotary shaker, 100 mg of sodium $[1,2\text{-}^{13}\text{C}_2]$ acetate was added to each flask, and the incubation was continued for an additional 4 days. The isolation procedure for $[1,2\text{-}^{13}\text{C}_2]$ acetate labeled rumbrin was essentially the same as previously reported¹⁾. Four mg of $[1,2\text{-}^{13}\text{C}_2]$ acetate labeled rumbrin was obtained from 1 liter culture broth.

References

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